

# Analysis and Prediction of Isotropic Mixing Magnetization Transfer Profiles in Three-Spin Topologies

Sarata C. Sahu<sup>1</sup>

Department of Chemical Sciences, Tata Institute of Fundamental Research, Homi Bhabha Road, Mumbai 400 005, India

Received May 30, 2000; revised July 24, 2000

**Isotropic mixing transfer functions ( $T_{kl}$ ) in three-spin systems typical of amino acids have been analyzed in order to develop simple rules for predicting transfer maxima/minima. For certain topologies, the intrinsically complex expressions describing the transfer functions reduce to compact forms which are easy to interpret and analyze. For other topologies where compactification is not possible, an analysis of the component functions of the  $T_{kl}$  reveals that only one or two components contribute significantly to the overall profile of the transfer function. As a result, simple rules of the thumb may be devised for reasonably accurate prediction of mixing times corresponding to local and global transfer maxima/minima, thereby facilitating mixing time optimization in TOCSY experiments.** © 2000 Academic Press

## INTRODUCTION

The description of isotropic mixing ( $I$ ) in multispin systems is essential for understanding and optimizing TOCSY ( $I, 2$ ) experiments. Unlike pulsed-free-precession experiments, analytical descriptions of multispin coupling topologies are possible only for very special situations ( $5-10$ ) and require numerical simulations in most cases ( $11-17$ ). Excellent analysis of isotropic mixing has appeared in Refs. (3) and (4).

Recently, Glaser and co-workers have derived analytical expressions for isotropic mixing transfer functions  $T_{kl}$  in a general three-spin (AMX) system ( $18$ ). It is obvious from the results that, except for very special cases, the  $T_{kl}$  are complicated sinusoidal functions of all relevant scalar couplings and the mixing time.

The rather complex nature of the transfer functions tends to reduce the utility of analytical expressions for predicting transfer maxima/minima, which is necessary for optimizing the mixing time in TOCSY experiments. Also, the complexity of the functions reveals very little insight into the dynamics of magnetization transfer in isotropic mixing. Any possibility of reducing these expressions to simpler forms or extracting their “key ingredients” to enhance their utility is always desirable.

In this paper, first we derive expressions for transfer functions in a general three-spin system using the relatively simple Liouville space framework, which is usually found in amino acid spin systems. Then, we consider topologies for which the expressions

reduce to a compact form. As discussed below, these situations correspond to commonly observed scenarios in amino acids. For these topologies, transfer maxima and corresponding mixing times can be predicted precisely. We then go on to show that even for other three-spin topologies, which do *not* belong to such “compact” categories, considerable simplification is possible if we analyze the *component* functions of a given  $T_{kl}$ , rather than the function as a whole. As a result, mixing times corresponding to transfer maxima/minima (in other words, the mixing time *profile*) can still be predicted with a reasonable degree of accuracy, which is important from a practical standpoint.

## ISOTROPIC MIXING IN A THREE-SPIN SYSTEM

For an arbitrary spin system  $\{I_1, I_2, I_3\}$ , the isotropic Hamiltonian is

$$H = 2\pi \sum_{k < l}^3 J_{kl} \mathbf{I}_k \cdot \mathbf{I}_l \quad [1]$$

Using the Liouville equation for the density operator  $\sigma$ , the time derivative for the expectation value of an operator  $A$  may be derived as

$$\dot{\sigma} = -i[H, \sigma] \quad [2]$$

$$\langle A \rangle = \text{Tr}(A\sigma) \quad [3]$$

$$\langle \dot{A} \rangle = -i \text{Tr}\{A[H, \sigma]\} \quad [4]$$

Using the property  $\text{Tr}\{A[B, C]\} = \text{Tr}\{[A, B]C\}$ , we have

$$\langle \dot{A} \rangle = -i \text{Tr}\{[A, H]\sigma\} \quad [5]$$

$$= -i\langle [A, H] \rangle \quad [6]$$

Using Eq. [6], the complete set of coupled differential equations may be written down for an appropriate  $n$ -dimensional operator basis  $\mathbf{A} \equiv \{A_k\}$ ,  $k = 1, n$ . An element  $A_k$  of this basis set is transformed by the commutator  $[A_k, H]$  into a linear combination of the basis operators:

$$[A_k, H] = \sum_l R_{kl} A_l \quad [7]$$

<sup>1</sup> Present address: Department of Biochemistry, University of California, Riverside, Riverside, CA 92521. E-mail: sarata@mail.ucr.edu.

Therefore, we may write Eq. [6] in matrix form,

$$\frac{d\mathbf{A}}{dt} = -i\mathbf{R}\mathbf{A}. \quad [8]$$

The isotropic Hamiltonian preserves coherence order. For a three-spin system, the following 15 normalized operators:  $I_x$ ,  $S_x$ ,  $Q_x$ ,  $2I_yS_z$ ,  $2I_zS_y$ ,  $2S_yQ_z$ ,  $2S_zQ_y$ ,  $2I_yQ_z$ ,  $2I_zQ_y$ ,  $4I_xS_yQ_y$ ,  $4I_xS_zQ_z$ ,  $4S_xI_yQ_y$ ,  $4S_xI_zQ_z$ ,  $4Q_xI_yS_y$ , and  $4Q_xI_zS_z$  represent an appropriate Liouville subspace. Differential equations can be written to describe the evolution of each of these operators. However, a suitable linear combination of these basis operators allows the set of 15 coupled differential equations to be partitioned into two sets of 9 and 6 equations each. The matrix equation for the nine-dimensional Liouville subspace is

$$\frac{d}{dt} \begin{pmatrix} \langle I_x \rangle - \langle S_x \rangle \\ \langle S_x \rangle - \langle Q_x \rangle \\ \langle I_x \rangle - \langle Q_x \rangle \\ IS \\ SQ \\ IQ \\ A - B \\ B - C \\ A - C \end{pmatrix} = \pi \begin{pmatrix} 0 & 0 & 0 & -2j_{12} & j_{23} & -j_{13} & 0 & 0 & 0 \\ 0 & 0 & 0 & j_{12} & -2j_{23} & -j_{13} & 0 & 0 & 0 \\ 0 & 0 & 0 & -j_{12} & -j_{23} & -2j_{13} & 0 & 0 & 0 \\ 2j_{12} & 0 & 0 & 0 & 0 & 0 & 0 & -j_{23} & j_{13} \\ 0 & 2j_{23} & 0 & 0 & 0 & 0 & -j_{12} & 0 & j_{13} \\ 0 & 0 & 2j_{13} & 0 & 0 & 0 & j_{12} & j_{23} & 0 \\ 0 & 0 & 0 & -j_{23} - j_{13} & 2j_{12} - j_{13} & j_{23} - 2j_{12} & 0 & 0 & 0 \\ 0 & 0 & 0 & 2j_{23} - j_{13} & -j_{12} - j_{13} & j_{12} - 2j_{23} & 0 & 0 & 0 \\ 0 & 0 & 0 & j_{23} - 2j_{13} & j_{12} - 2j_{13} & -j_{12} - j_{23} & 0 & 0 & 0 \end{pmatrix} \cdot \begin{pmatrix} \langle I_x \rangle - \langle S_x \rangle \\ \langle S_x \rangle - \langle Q_x \rangle \\ \langle I_x \rangle - \langle Q_x \rangle \\ IS \\ SQ \\ IQ \\ A - B \\ B - C \\ A - C \end{pmatrix}, \quad [9]$$

where  $j_{12}$ ,  $j_{13}$  and  $j_{23}$  are the coupling constants between I, S; I, Q; S, Q, respectively, and

$$A = \langle I_x S_y Q_y \rangle + \langle I_x S_z Q_z \rangle \quad [10]$$

$$B = \langle S_x I_y Q_y \rangle + \langle S_x I_z Q_z \rangle \quad [11]$$

$$C = \langle Q_x I_y S_y \rangle + \langle Q_x I_z S_z \rangle \quad [12]$$

$$IS = \langle I_y S_z \rangle - \langle I_z S_y \rangle \quad [13]$$

$$SQ = \langle S_y Q_z \rangle - \langle S_z Q_y \rangle \quad [14]$$

$$IQ = \langle I_y Q_z \rangle - \langle I_z Q_y \rangle, \quad [15]$$

whereas the matrix equation for the six-dimensional subspace is

$$\frac{d}{dt} \begin{pmatrix} IS^+ \\ SQ^+ \\ IQ^+ \\ A^- \\ B^- \\ C^- \end{pmatrix} = \pi \begin{pmatrix} 0 & 0 & 0 & -j_{13} & -j_{23} & j_{23} + j_{13} \\ 0 & 0 & 0 & -j_{12} & j_{12} + j_{23} & -j_{23} \\ 0 & 0 & 0 & j_{12} + j_{13} & -j_{12} & -j_{13} \\ j_{13} & j_{12} & -j_{12} - j_{13} & 0 & 0 & 0 \\ j_{23} & -j_{12} - j_{23} & j_{12} & 0 & 0 & 0 \\ -j_{23} - j_{13} & j_{23} & j_{13} & 0 & 0 & 0 \end{pmatrix} \cdot \begin{pmatrix} IS^+ \\ SQ^+ \\ IQ^+ \\ A^- \\ B^- \\ C^- \end{pmatrix}, \quad [16]$$

$$A^- = \langle I_x S_y Q_y \rangle - \langle I_x S_z Q_z \rangle \quad [17]$$

$$B^- = \langle S_x I_y Q_y \rangle - \langle S_x I_z Q_z \rangle \quad [18]$$

$$C^- = \langle Q_x I_y S_y \rangle - \langle Q_x I_z S_z \rangle \quad [19]$$

$$IS^+ = \langle I_y S_z \rangle + \langle I_z S_y \rangle \quad [20]$$

$$SQ^+ = \langle S_y Q_z \rangle + \langle S_z Q_y \rangle \quad [21]$$

$$IQ^+ = \langle I_y Q_z \rangle + \langle I_z Q_y \rangle. \quad [22]$$

Interestingly, both sets of equations are amenable to analytical solutions using standard methods, such as Laplace transforms or matrix diagonalization. We found the Laplace transform method more easy to implement on *Mathematica* version 2.2. While the procedure is standard textbook material, it is

outlined below for completeness. Taking Laplace transforms on both sides of Eq. [6] we have

$$s\mathcal{L}_A - \mathbf{A}(0) = -i\mathbf{R}\mathcal{L}_A, \quad [23]$$

$\mathcal{L}_A$  being the Laplace transform of  $\mathbf{A}$ . After rearranging Eq. [23], we have

$$(s\mathbf{E} + i\mathbf{R})\mathcal{L}_A = \mathbf{A}(0), \quad [24]$$

where  $\mathbf{E}$  is the identity operator. Denoting  $\mathbf{U} = s\mathbf{E} + i\mathbf{R}$ , we have

**TABLE 1**  
**Expressions of Transfer Functions for Isotropic Mixing in a General Three-Spin System**

$$\begin{aligned}\langle T \rangle_{11} &= \frac{1}{3} + \frac{(j_{12} + 2j_{23} + j_{13})^2}{18D^2} + \frac{(j_{13} - j_{12})^2}{6D^2} \cos \lambda_1 t + \frac{2D - j_{12} + 2j_{23} - j_{13}}{9D} \cos \lambda_2 t + \frac{2D + j_{12} - 2j_{23} + j_{13}}{9D} \cos \lambda_3 t \\ \langle T \rangle_{12} &= \frac{1}{3} + \frac{(j_{12} + j_{23} - 2j_{13}) \cdot (j_{12} - 2j_{23} + j_{13})}{18D^2} + \frac{(j_{12} - j_{23}) \cdot (j_{13} - j_{12})}{6D^2} \cos \lambda_1 t + \frac{2j_{12} - j_{23} - j_{13} - D}{9D} \cos \lambda_2 t + \frac{-2j_{12} + j_{23} + j_{13} - D}{9D} \cos \lambda_3 t \\ \langle T \rangle_{13} &= \frac{1}{3} + \frac{(j_{12} + 2j_{23} + j_{13}) \cdot (-2j_{12} + j_{23} + j_{13})}{18D^2} + \frac{(j_{23} - j_{13}) \cdot (j_{13} - j_{12})}{6D^2} \cos \lambda_1 t + \frac{-j_{12} - j_{23} + 2j_{13} - D}{9D} \cos \lambda_2 t + \frac{j_{12} + j_{23} - 2j_{13} - D}{9D} \cos \lambda_3 t\end{aligned}$$

Note.  $d = (j_{12} + j_{23} + j_{13})$ ;  $D = (j_{12}^2 + j_{23}^2 + j_{13}^2 - (j_{12}j_{23} + j_{23}j_{13} + j_{12}j_{13}))^{1/2}$ ;  $\lambda_1 = 2\pi D$ ;  $\lambda_2 = \pi(d - D)$ ;  $\lambda_3 = \pi(d + D)$ .

$$\mathcal{L}_A = \mathbf{U}^{-1} \mathbf{A}(0) \quad [25]$$

$$f_j(\tau_m) = a_j \cdot \cos(2\pi \nu_j \tau_m), \quad [29]$$

and, consequently,

$$\mathbf{A}(t) = \mathbf{V} \mathbf{A}(0) \quad [26]$$

$$\mathbf{V} = \mathcal{L}^{-1} \mathbf{U}^{-1}. \quad [27]$$

Here we present results obtained for the usually encountered initial condition where the magnetization resides in-phase on one of the spins. This results in only the nine-dimensional subspace being relevant for the solutions. The expressions for transfer functions pertaining to in-phase and two-spin anti-phase operators are listed in Tables 1 and 2, respectively. Using accepted conventions, we use the symbol  $T_{kl}$  to denote the  $k \rightarrow l$  transfer function which, in the Liouville space formalism, corresponds to the value of  $\langle A_l \rangle(t)$  for the initial condition  $\langle A_k \rangle(0) = 1$ , others zero. From the expression for  $T_{kl}$ , other transfer functions (e.g.,  $T_{lm}$ ) may be obtained by simple permutation of the spin identities.

The three-spin topology found typically in biomolecules consists of two vicinal and one geminal coupling (Fig. 1), usually an  $H_\alpha$ - $H_{\beta 1}$ - $H_{\beta 2}$  system in Phe, Tyr, Trp, His, Asp, Asn, Cys, and Ser. In this work, we refer to the vicinal  $J_{\alpha\beta 1,2}$  as  $\nu_1$  ( $j_{12}$ ) and  $\nu_2$  ( $j_{13}$ ), respectively, and the geminal  $J_{\beta 1\beta 2}$  coupling as  $g$  ( $j_{23}$ ), with  $\{\nu_1, \nu_2\} \leq |g|$ . The validity of the conclusions made in this paper has been tested for  $0 \leq \{\nu_1, \nu_2\} \leq 15$  Hz and  $-16 < g < -12$ , which are the usually observed ranges of these couplings in amino acids (20–23). For the simulations shown in the paper, we have assumed a value of  $-15$  Hz for  $g$ . Finally, we have concentrated only on “cross-peak intensities,” i.e.,  $T_{kl}$ ,  $k \neq l$ .

### ANALYSIS OF TOPOLOGIES YIELDING COMPACT SOLUTIONS

In general, the total transfer function  $T_{kl}(\tau_m)$  for an  $n$ -spin system is given by (4)

$$T_{kl}(\tau_m) = a_0 + \sum_{j=1}^N f_j(\tau_m) \quad [28]$$

where  $N = 1$  and 3 for two- and three-spin systems, respectively, and  $\nu_j$  are derived from differences of eigenvalues of  $H$ . The number  $N$  grows rapidly with the number of spins in the network (e.g.,  $N = 12$  for a four-spin system) (19).

Since  $T_{kl}$  is a superposition of several trigonometric functions, it is not trivial to calculate maxima and minima even if all the  $a_j$  and  $\nu_j$  are known. However, for certain spin topologies, the individual functions  $f_j$  seldom contribute equally to  $T_{kl}$ , which then becomes far more tractable for analysis.

From Eq. [29], the vicinal transfer function  $T_{12}$  for a three-spin system may be written as

$$T_{12}(\tau_m) = a_0 + f_1(\tau_m) + f_2(\tau_m) + f_3(\tau_m). \quad [30]$$

Based on the results of Table 1 and with  $\nu_{1,2}$  as defined above, the coefficients  $a_j$  and functions  $f_j$  are

$$a_0 = 1/3 + (\nu_1 + \nu_2 - 2g)(\nu_1 - 2\nu_2 + g)/(18D^2) \quad [31]$$

$$f_1 = a_1 \cdot \cos(2\pi D \tau_m) \quad [32]$$

$$f_2 = a_2 \cdot \cos[\pi(d - D)\tau_m] \quad [33]$$

$$f_3 = a_3 \cdot \cos[\pi(d + D)\tau_m] \quad [34]$$

$$a_1 = \frac{(\nu_1 - \nu_2)(g - \nu_1)}{6D^2} \quad [35]$$

$$a_2 = \frac{(2\nu_1 - \nu_2 - g - D)}{9D} \quad [36]$$

$$a_3 = \frac{-(2\nu_1 - \nu_2 - g + D)}{9D} \quad [37]$$

$$d = g + \nu_1 + \nu_2 \quad [38]$$

$$D = [g^2 + \nu_1^2 + \nu_2^2 - (g\nu_1 + g\nu_2 + \nu_1\nu_2)]^{1/2}. \quad [39]$$

$T_{13}$  and  $T_{23}$  are obtained easily from Eqs. [31]–[39], by appropriately permuting the identities of the spins. Two situations where  $T_{12}$  takes a simple form are the following.

**TABLE 2**  
**Expressions for Transfer Coefficients for Transfer from  $I_x \rightarrow$  Different Antiphase Terms**  
**for Isotropic Mixing in a General Three-Spin System**

---


$$\langle 2I_y S_z \rangle = -\langle 2I_z S_y \rangle = \frac{((j_{12} - j_{13})\sin \lambda_1 + (-j_{12} + j_{23} + D)\sin \lambda_2 + (j_{12} - j_{23} + D)\sin \lambda_3)}{6D}$$

$$\langle 2S_y Q_z \rangle = -\langle 2S_z Q_y \rangle = \frac{(j_{12} - j_{13}) \cdot (\sin \lambda_1 + \sin \lambda_2 - \sin \lambda_3)}{6D}$$

$$\langle 2I_y Q_z \rangle = -\langle 2I_z Q_y \rangle = \frac{((j_{13} - j_{12})\sin \lambda_1 + (j_{23} - j_{13} + D)\sin \lambda_2 + (-j_{23} + j_{13} + D)\sin \lambda_3)}{6D}$$

$$\langle I_x S_z Q_z \rangle = \langle I_x S_y Q_y \rangle = -\frac{(j_{12} - 2j_{23} + j_{13})^2}{18D^2} - \frac{(j_{13} - j_{12})^2}{6D^2} \cos \lambda_1 + \frac{(2D - j_{12} + 2j_{23} - j_{13})}{18D} \cos \lambda_2 + \frac{(2D + j_{12} - 2j_{23} + j_{13})}{18D} \cos \lambda_3$$

$$\langle S_x I_z Q_z \rangle = \langle S_x I_y Q_y \rangle$$

$$= \frac{(j_{12} - 2j_{23} + j_{13}) \cdot (-j_{12} - j_{23} + 2j_{13})}{18D^2} + \frac{(j_{23} - j_{12}) \cdot (j_{13} - j_{12})}{6D^2} \cos \lambda_1 + \frac{(2j_{12} - j_{23} - j_{13} - D)}{18D} \cos \lambda_2 + \frac{(-2j_{12} + j_{23} + j_{13} - D)}{18D} \cos \lambda_3$$

$$\langle Q_x I_z S_z \rangle = \langle Q_x I_y S_y \rangle$$

$$= \frac{(2j_{12} - j_{23} - j_{13}) \cdot (j_{12} - 2j_{23} + j_{13})}{18D^2} + \frac{(j_{12} - j_{13}) \cdot (j_{23} - j_{13})}{6D^2} \cos \lambda_1 + \frac{(-j_{12} - j_{23} + 2j_{13} - D)}{18D} \cos \lambda_2 + \frac{(j_{12} + j_{23} - 2j_{13} - D)}{18D} \cos \lambda_3$$


---

Note.  $d$ ,  $D$ ,  $\lambda_1$ ,  $\lambda_2$ , and  $\lambda_3$  are as defined for Table 1.

(a)  $v_1 = v_2 = v$

This has been dealt with in detail by Glaser (9) and corresponds to gauche-gauche or rotationally averaged conformations in amino acid side chains.  $T_{12} = T_{13}$  takes the form

$$T_{12} = T_{13} = \frac{2}{9} [1 - 2 \cos(3\pi vt)]. \quad [40]$$

As noted by Glaser (9), the transfer is independent of  $g$ . Transfer maxima ( $T_{12}^{\max} = T_{13}^{\max} = 33\%$ ) appear at odd multiples of  $t = 1/(3v)$ .

(b)  $d \sim 0$ , or  $d/D \ll 1$ , i.e.,  $g \approx -(v_1 + v_2)$

This is not an uncommon situation in amino acids, typical examples being the *trans*-gauche conformation ( $v_1 = 3$ ,  $v_2 =$

12,  $g = -15$ ) or ( $v_1 = 6$ ,  $v_2 = 9$ ,  $g = -15$ ) (20). Considering the situation where  $d$  is exactly zero, the expressions for  $T_{12}$  and  $T_{13}$  become

$$T_{12} = \frac{1}{3} - \frac{v_2(v_1 + v_2)}{2D^2} + \frac{(2v_1 + v_2)(v_2 - v_1)}{6D^2} \times \cos 2\pi Dt - \frac{2}{9} \cos \pi Dt \quad [41]$$

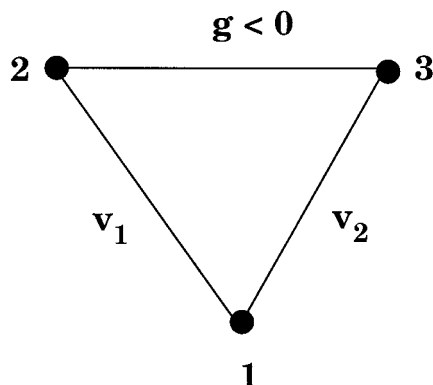
$$T_{13} = \frac{1}{3} - \frac{v_1(v_2 + v_1)}{2D^2} - \frac{(2v_2 + v_1)(v_1 - v_2)}{6D^2} \times \cos 2\pi Dt - \frac{2}{9} \cos \pi Dt \quad [42]$$

$$D = [3(v_1^2 + v_2^2 + v_1 v_2)]^{1/2}. \quad [43]$$

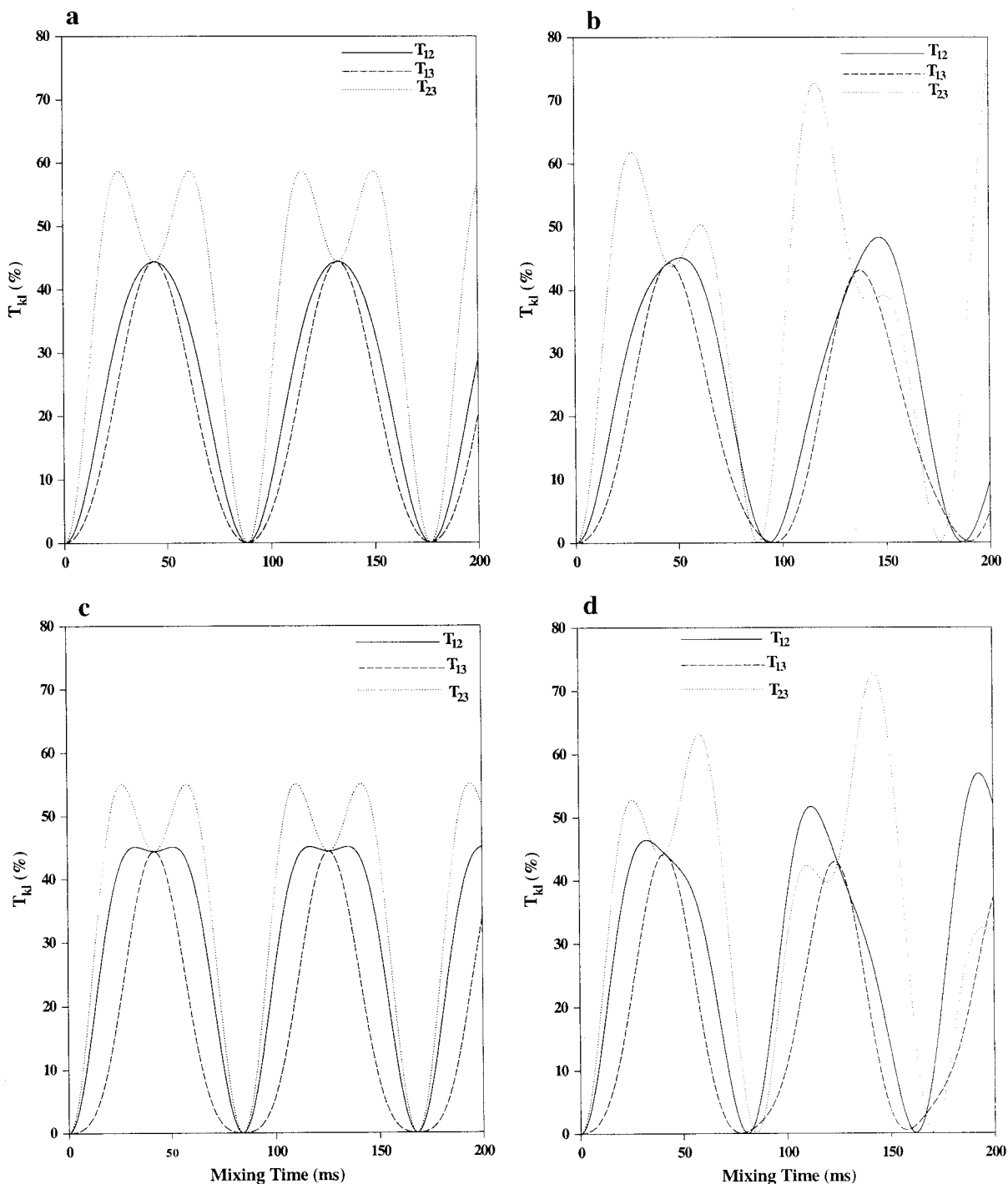
By setting the time derivatives to zero, it is easily shown that for  $z = (v_1/v_2) < (1 + \sqrt{3})$ , extrema correspond to

$$\sin \pi D \tau_m^{\max} = 0 \quad z = v_1/v_2 < (1 + \sqrt{3}). \quad [44]$$

The interesting point to note here is that for both  $T_{12}$  and  $T_{13}$ , peak (maxima or in some cases local minima) appear at the same mixing time ( $1/D$ ), with identical efficiencies of 44.4%, regardless of the relative magnitudes of  $v_1$  and  $v_2$ . As a result, both of the cross peaks can be maximized at the same mixing time.  $T_{23}$  also has the same value (44.4%) at  $t = 1/D$ , although it does not correspond to a maximum. This is illustrated in Fig. 2a, which shows the three transfer functions, assuming  $v_1 = 9.0$  and  $v_2 = 6.0$  Hz.  $T_{12}$ , associated with the larger coupling  $v_1$ , builds up



**FIG. 1.** A general three-spin system typically seen in biomolecules. Here  $v_1$  and  $v_2$  are vicinal and  $g$  is the geminal coupling constant.



**FIG. 2.** Mixing time dependence of transfer functions at different values of  $v_1$  and  $v_2$ : (a)  $v_1 = 9.0$  Hz and  $v_2 = 6.0$  Hz, (b)  $v_1 = 9.0$  Hz and  $v_2 = 4.0$  Hz, (c)  $v_1 = 12.0$  Hz and  $v_2 = 3.0$  Hz, (d)  $v_1 = 12.0$  Hz and  $v_2 = 3.5$  Hz. The value of  $g$  is  $-15$  Hz in all cases.

rapidly in the beginning, but the slower function  $T_{13}$  catches up steeply in the vicinity of  $t = 1/D$ . In situations where  $d \neq 0$  but  $d \ll D$ , the maxima of  $T_{12}$  and  $T_{13}$  are marginally offset from each other, but the point of intersection lies very close to the larger/smaller of the two peaks, as shown in Fig. 2b ( $v_1 = 9.0$ ,  $v_2 = 4.0$ ,  $d/D = -0.09$ ). In addition,  $T_{23}$  develops multiple maxima, which exceed that of  $T_{12}$  and  $T_{13}$ .

For  $z \geq (1 + \sqrt{3})$ , the condition for extrema is given by

$$\cos \pi D \tau_m^{\max} = \frac{v_1^2 + v_2^2 + v_1 v_2}{(v_2 - v_1)(2v_1 + v_2)} \quad z \geq (1 + \sqrt{3}). \quad [45]$$

An example of the mixing time profile of a topology in this

category ( $v_1 = 12$ ,  $v_2 = 3$ ) is shown in Fig. 2c.  $T_{12}$  displays multiple maxima at

$$\tau_m^{\max} = \frac{1}{\pi D} \left[ 2n\pi \pm \arccos \left\{ \frac{v_1^2 + v_2^2 + v_1 v_2}{(v_2 - v_1)(2v_1 + v_2)} \right\} \right]$$

$$n = 1, 2, \dots \quad [46]$$

Since the condition in Eq. [45] requires one vicinal coupling to be almost four times the other, it occurs only for classic *trans-gauche* conformations. The  $T_{12}$  and  $T_{13}$  curves meet at  $1/D$ , which is a  $T_{12}$  local minimum sandwiched between the two symmetrical  $T_{12}$  maxima. If  $d$  deviates slightly from zero as shown in Fig. 2d ( $v_1 = 12$ ,  $v_2 = 3.5$ ,  $d/D = 0.02$ ), the  $T_{12}$  profile becomes skewed, but the overall profile remains similar.

### ANALYSIS OF TOPOLOGIES WITHOUT COMPACT SOLUTIONS

For topologies which do not belong to either of the above two categories, it is still possible to predict the profile of  $T_{kl}$  vs  $\tau_m$  with reasonable accuracy, over the vicinal coupling constant range  $0 \leq \{v_1, v_2\} \leq 15$  Hz. Figures 3a–3d show  $T_{12}$  and  $T_{13}$  for two topologies for which  $v_1 \neq v_2$  and neither is  $v_1 + v_2 \sim |g|$ . In each of the plots, the  $T_{kl}$ ,  $f_1$ ,  $f_3$ , and  $Q_{kl} = f_1 + f_3$  are shown as a function of mixing time. It is easily seen that the sum ( $Q_{kl}$ ) of the fastest frequency component ( $f_1$ ) and the largest amplitude component ( $f_3$ ) reproduce the overall profile of  $T_{kl}$  very closely. While it is easily demonstrated from the definitions of  $d$  and  $D$  that  $f_1$  is the most rapidly oscillating component for all  $v_1$ ,  $v_2$ , and  $g$ ,  $a_3$  is not always the largest coefficient. However, numerical calculations show that  $a_3$  is indeed the largest coefficient in the range  $0 \leq \{v_1, v_2\} \leq 15$ . Although a value of  $g = -15$  Hz has been used in Fig. 3, the validity of the result has been verified over the range  $12 < |g| < 16$ .

Essentially, a knowledge of the extrema of  $f_1$  (at multiples of  $1/D$ ) and  $f_3$  (at multiples of  $2/(d + D)$ ) is a good indicator of the profile of the overall function. Two different situations may be considered, as follows.

$v_1 + v_2 < |g|$ . Each local maximum or minimum of  $T_{kl}$  usually coincides with corresponding extrema of  $f_1$ . If  $v_1 < v_2$  (Fig. 3a),  $a_1$  is positive (Eq. [35]) and, therefore,  $f_1$  maxima occur at  $t_{f_1}^{\max} = n/D$ ,  $n = 1, 2, \dots$ . If  $v_1 > v_2$ ,  $a_1$  is negative and, therefore,  $f_1$  maxima appear at  $t_{f_1}^{\max} = (2n - 1)/(2D)$ ,  $n = 1, 2, \dots$ . For  $v_1 < v_2/2$ ,  $a_3$  is obviously negative (Eq. [37]). Otherwise,  $a_3$  becomes positive only when  $(v_2 - 2v_1) > (|g| + D)$ . Both  $|g|$  and  $D$  are large quantities and, therefore, this condition requires  $v_2$  to be abnormally large, which is never true for any realistic values of proton coupling constants. As a result,  $a_3$  may be considered negative and, therefore,  $f_3$  maxima appear at  $t_{f_3}^{\max} = (2n - 1)/(d + D)$ .

“Global” maxima in  $T_{kl}$  arise when  $f_1$  and  $f_3$  are maximally

in-phase, i.e., when maxima of  $f_1$  coincide with that of  $f_3$ . When  $f_1$  and  $f_3$  are anti-phase, i.e., an  $f_1$  maximum coincides with an  $f_3$  minimum, the situation corresponds to a global *minimum* in  $T_{kl}$ , since  $a_3$  is always the more dominant coefficient. From a practical viewpoint, these observations may be encapsulated into the following protocol: Tabulate all values of  $t_{f_1}^{\max}$ ,  $t_{f_1}^{\min}$ ,  $t_{f_3}^{\max}$ , and  $t_{f_3}^{\min}$  in a given mixing time range. In general, a  $t_{f_1}^{\max}$  corresponds to a local maximum in the  $T_{kl}$ . If the  $t_{f_1}^{\max}$  also happens to be coincident with or appears close to a  $t_{f_3}^{\max}$ , the situation corresponds to a global maximum. On the other hand, if  $t_{f_1}^{\max}/t_{f_1}^{\min}$  lies close to a  $t_{f_3}^{\min}$ , a global minimum results.

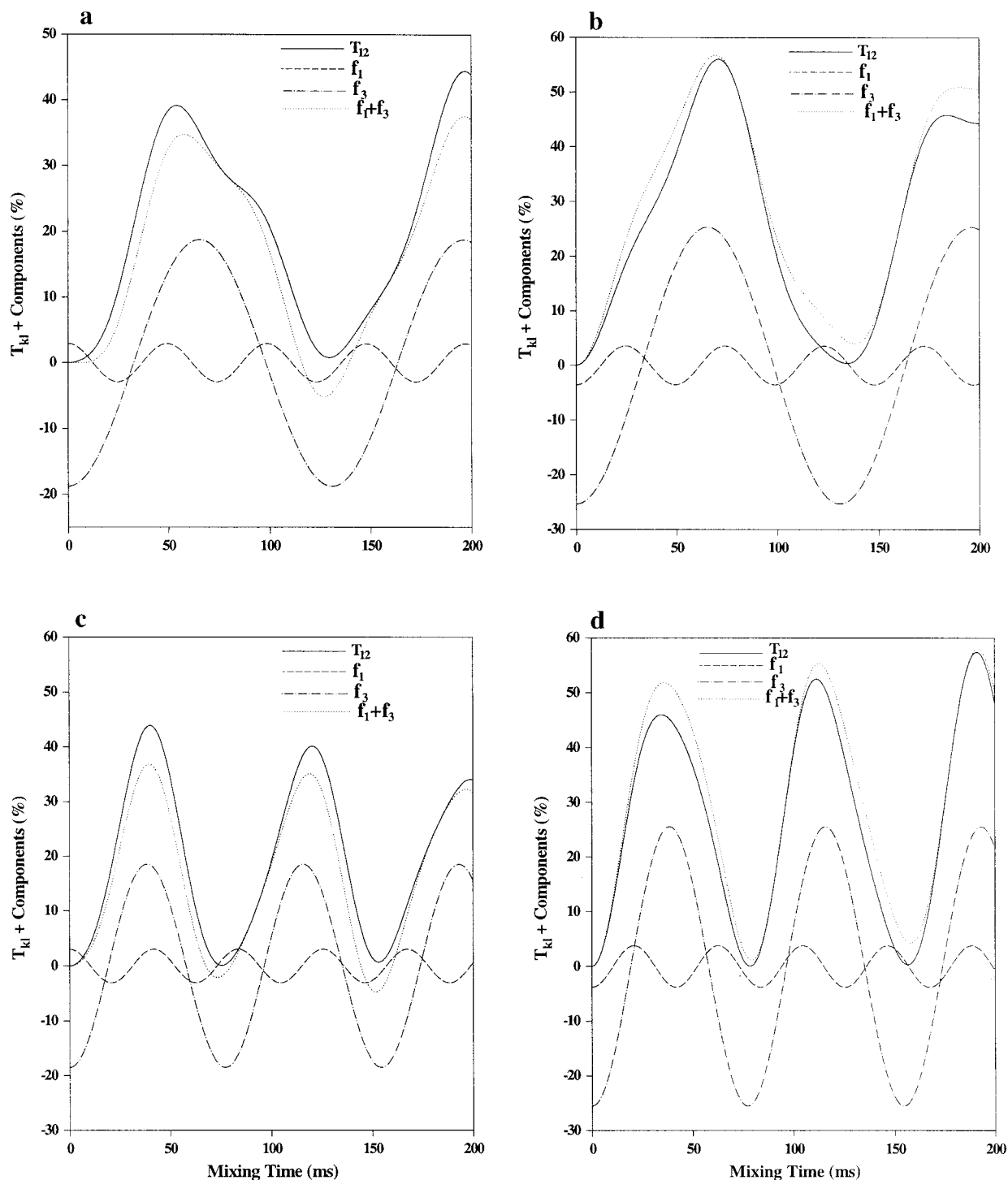
When  $v_1 + v_2 > |g|$  (Figs. 3c and 3d), the denominator of  $a_1$  ( $6D^2$ ) becomes large and, therefore,  $a_1$  is small. The  $T_{kl}$  profile is, therefore, almost completely dominated by  $f_3$ . Maxima and minima in  $T_{kl}$  appear at mixing times close to  $t_{f_3}^{\max} = (2n - 1)/(d + D)$ . The  $f_3$  maxima are broad, typically spanning 10 ms. As a result, differences between the value of  $T_{kl}$  at  $t_{f_3}^{\max}$  and the actual  $T_{kl}^{\max}$  is very small (Figs. 3c and 3d) and largely insensitive to the relative magnitudes of  $v_1$  and  $v_2$ .

Combining the compact solutions with the “rules” developed in this section, mixing times corresponding to transfer function maxima/minima may be obtained largely from a knowledge of  $d$  and  $D$  alone.

### CONCLUSIONS

The isotropic mixing transfer functions in three-spin systems are obtained using an alternative approach to that of Glaser *et al.* (18) and analyzed these functions in order to extract their information content in a simple manner. For certain spin topologies typical of amino acids, the intrinsically complex functions reduce to compact forms which are easy to use for predicting transfer maxima/minima. An interesting result occurs when  $v_1 + v_2 \sim |g|$ , wherein both the transfer functions  $T_{12}$  and  $T_{13}$  peak at the same mixing time with the same value, regardless of the relative magnitudes of  $v_1$  and  $v_2$ . For other topologies where compactification is not possible, in the normally observed coupling constant ranges of  $0 \leq \{v_1, v_2\} \leq 15$  Hz and  $-16 \leq g \leq -12$ , an analysis of the component functions of the  $T_{kl}$  reveals that only one or two components contribute significantly to the overall profile of the transfer function. This leads to simple rules of the thumb for reasonably accurate prediction of mixing times corresponding to local and global transfer maxima/minima and thereby facilitates mixing time optimization in experiments. The transfer functions presented in this paper correspond to the ideal isotropic mixing case, where effect relaxation is neglected, which often leads to an additional damping of the experimental transfer function. In practice, relaxation effects are more severe at longer mixing times and need to be accounted for. However, for small to medium-size molecules in which proton TOCSY experiments are most relevant, these expressions are likely to be applicable up to 100 ms of mixing time, depending on





**FIG. 3.** Mixing time dependence of transfer functions and their various components at different values of  $v_1$  and  $v_2$ : (a)  $v_1 = 3.0$  Hz and  $v_2 = 7.0$  Hz, (b)  $v_1 = 7.0$  Hz and  $v_2 = 3.0$  Hz, (c)  $v_1 = 6.0$  Hz and  $v_2 = 11.0$  Hz, (d)  $v_1 = 11.0$  Hz and  $v_2 = 6.0$  Hz. The value of  $g$  is always taken as  $-15$  Hz.

the molecular weight. While this analysis has been restricted to the smallest of multispin systems, investigation of isotropic mixing in four-spin systems has indicated that even when the number of component functions increases signif-

icantly (12 for 4-spin systems), about 6 of 12 harmonic terms contribute substantially to the overall  $T_{kl}$  (19). This holds out promise for similar analyses of more complex spin systems.

## ACKNOWLEDGMENT

I thank Dr. Ananya Majumdar for his constant support, suggestions, and many useful discussions.

## REFERENCES

1. L. Braunschweiler and R. R. Ernst, Coherence transfer by isotropic mixing: Application of proton correlation spectroscopy, *J. Magn. Reson.* **53**, 521–528 (1983).
2. A. Bax and D. G. Davis, MLEV-17-based two-dimensional homonuclear magnetization transfer spectroscopy, *J. Magn. Reson.* **65**, 355–360 (1985).
3. S. J. Glaser and G. P. Drobny, in "Advances in Magnetic Resonance," (J. S. Waugh, Ed.), Vol. 14, p. 35–58, Academic Press, San Diego.
4. S. J. Glaser and J. J. Quant, in "Advances in Magnetic and Optical Resonance," (W. S. Warren, Ed.), Vol. 19, pp. 59–252, Academic Press, San Diego.
5. N. Chandrakumar and S. Subramanian, Some aspects of coherence transfer by isotropic mixing, *J. Magn. Reson.* **62**, 346–349 (1985).
6. N. Chandrakumar, G. V. Visalakshi, D. Ramaswamy, and S. Subramanian, Analysis of collective modes in some  $A_M X_N$  systems, *J. Magn. Reson.* **67**, 307–318 (1986).
7. G. V. Visalakshi and N. Chandrakumar, Automated generation of commutators algebra for NMR problems, *J. Magn. Reson.* **75**, 1–8 (1987).
8. J. Listerud, S. J. Glaser, and G. P. Drobny, Symmetry and isotropic coherence transfer; II. Multi spin calculations using a young tableaux formulation, *Mol. Phys.* **78**, 629–658 (1993).
9. S. J. Glaser, Coupling topology dependence of polarization-transfer efficiency in TOCSY and TACSU experiments, *J. Magn. Reson. A* **104**, 283–301 (1993).
10. A. Majumdar, Analytical expressions for isotropic mixing in three- and four-spin topologies in  $^{13}\text{C}$  systems, *J. Magn. Reson. A* **121**, 121–126 (1996).
11. M. Rance, Sign reversal of resonances via isotropic mixing in NMR spectroscopy, *Chem. Phys. Lett.* **154**, 242–247 (1989).
12. M. L. Remerowski, S. J. Glaser, and G. P. Drobny, A theoretical study of coherence transfer by isotropic mixing: Calculation of pulse sequence performance for systems of biological interest, *Mol. Phys.* **68**, 1191–1218 (1989).
13. J. Cavanagh, W. J. Chazin, and M. Rance, The time dependence of coherence transfer in homonuclear isotropic mixing experiments, *J. Magn. Reson.* **87**, 110–131 (1990).
14. A. Bax, G. M. Clore, and A. M. Gronenborn,  $^1\text{H}$ – $^{13}\text{C}$  correlation via isotropic mixing of  $^{13}\text{C}$  magnetization—A new three-dimensional approach for assigning  $^1\text{H}$  and  $^{13}\text{C}$  spectra of  $^{13}\text{C}$ -enriched proteins, *J. Magn. Reson.* **88**, 425–431 (1990).
15. H. S. Eaton, S. W. Fesik, S. J. Glaser, and G. P. Drobny, Time dependence of  $^{13}\text{C}$ – $^{13}\text{C}$  magnetization transfer in isotropic mixing experiments involving amino acid spin systems, *J. Magn. Reson.* **90**, 452–463 (1990).
16. S. S. Wijmenga, H. A. Heus, B. Werten, G. A. van der Marel, J. H. van Boom, and C. W. Hilbers, Assignment strategies and analysis of cross-peak patterns and intensities in the three-dimensional homonuclear TOCSY-NOESY of RNA, *J. Magn. Reson. B* **103**, 134 (1994).
17. P. Khandelwal, A. Majumdar, and R. V. Hosur, Mixing time dependence of cross peak intensities in  $^{13}\text{C}$  resolved  $^1\text{H}$ – $^1\text{H}$  TOCSY spectra of proteins, *Proc. Natl. Acad. Sci. India* **66(A)**, Special Issue, 1996.
18. O. Schedletsky and S. J. Glaser, Analytical coherence-transfer functions for the general AMX spin system under isotropic mixing, *J. Magn. Reson. A* **123**, 174–180 (1996).
19. B. Luy, O. Schedletsky, and S. J. Glaser, Analytical polarization transfer functions for four coupled spins  $\frac{1}{2}$  under isotropic mixing conditions, *J. Magn. Reson. A* **138**, 19–27 (1999).
20. H. Widmer and K. Wuthrich, Simulated two-dimensional NMR cross-peak fine structures for  $^1\text{H}$  spin systems in polypeptides and polydeoxynucleotides, *J. Magn. Reson.* **74**, 316–336 (1987).
21. K. Bartik and C. Redfield, A method for the estimation of  $^1\text{H}$  torsion angles in proteins, *J. Biomol. NMR* **3**, 415–428 (1993).
22. M. Cai, J. Liu, Y. Gong, and R. Krishnamoorthi, A practical method for stereospecific assignments of  $\gamma$ - and  $\delta$ -methylene hydrogens via estimation of vicinal  $^1\text{H}$ – $^1\text{H}$  coupling constants, *J. Magn. Reson. B* **107**, 172–178 (1995).
23. K. L. Constantine, M. S. Friedrichs, and L. Mueller, Simple approaches for estimating vicinal  $^1\text{H}$ – $^1\text{H}$  coupling constants and for obtaining stereospecific resonance assignments in leucine side chains, *J. Magn. Reson. B* **104**, 62–68 (1994).
24. J. Briandt and R. R. Ernst, Sensitivity comparison of two-dimensional correlation spectroscopy in the laboratory frame and in the rotating frame, *J. Magn. Reson. A* **104**, 54–62 (1993).